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# Nutritional and Antioxidant Properties of Yoghurt Enriched with Pineapple and Orange Pomace

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### **Abstract**

The present study aimed to evaluate the physiochemical, and nutritional properties of the yoghurt enriched with pineapple and orange pomace. Five yoghurt samples were prepared using skimmed milk, orange pomace, and pineapple pomace blends. The physicochemical properties (pH, total titratable acid, free fatty acidity, total soluble solid, and brix), anti-nutrient properties (saponins, oxalate, phytate, tannins, and alkaloids), and antioxidant properties [ferric reducing antioxidant properties (FRAP), 2, 2'-azino-bis (3-ethylbenthiazoline-6-sulphonic acid (ABTS), 1, 1- diphenyl-2-picryhydrazyl (DPPH), phenol and iron chelate (Fe<sup>2+</sup>) of the samples were evaluated. The study showed that pineapple and orange pomaces affected the pH and viscosity of the samples. The mean and standard deviation of the samples were determined and a One-Way Analysis of Variance (ANOVA). There were significant increase (p≤0.05) in the free fatty acid (9.10-12.00%), total soluble solid (6.30 10.50%), total titratable acid (8.50-14.93%), and brix (11.50-17.00%) compared with the control. The anti-nutritional factors were within acceptable standards and sample YOB (75.04% Skimmed Yoghurt, 22.65% Pineapple Pomace and 2.87% Orange Pomace) had the best antinutritional and antioxidants properties. Therefore, Pineapple and Orange pomace can be used as a functional ingredient for the development of Skimmed Milk yoghurt.

**Keywords:** Yoghurt, pomace, physicochemical, nutritional properties, functional drink

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## Introduction

Yoghurt is formed by fermenting milk with *Streptococcus thermophilus* and *Lactobacillus delbrueckii subsp bulgaricus*, resulting in its distinct flavor and texture (Arijit *et al.*, 2020). According to Rodriguez *et al.* (2006), adding pomace powder to yoghurt enhances its physicochemical properties, nutritional qualities, and functional benefits such as swelling, water retention, emulsification, and textural enhancement. Enriched yogurt with these advantages can be a healthier choice for consumers compared to conventional yogurts (Olabiran *et al.*, 2023: Noreen *et al.*, 2019).

Pomaces are solid waste products of fruits rich in dietary fiber, they are pulpy residue remaining after extracting the juice of a fruit (Hussein *et al.*, 2015). They can be obtained from fruits after processing for juice, pulps, jams, and jellies (Awolu *et al.*, 2016). Recommended globally for its health benefits pomace fiber possesses unique functional properties Utilizing by-products from fruit and vegetable processing is a crucial and challenging task globally (Homayouni *et al.*, 2013: Tran, 2006). Pineapple pomace contains high levels of dietary fiber, vitamins, and minerals (Wang *et al.*, 2002) Dietary fiber is mainly incorporated into cereal-based products, particularly bakery products, meat and milk products (Dhingra *et al.*,

modifying the texture, taste, and shelf life of food products, such as water-holding capacity, gel-forming ability, fat mimetic properties, thickening characteristics, and anti-staling benefits (Charoenthaikij *et al.*, 2016: Sabanis *et al.*, 2009). Citrus pomaces provide a rich source of bioactive ingredients with excellent technological and nutritional properties (Awolu *et al.*, 2020). Their affordability and versatility make them ideal components or additives in food products and are also cost-effective (Sánchez *et al.*, 2017). Pineapple (*Ananas comosus*) is mainly consumed fresh or processed as canned fruit the increasing production of processed pineapple and citrus juice results in significant waste and by-products. Utilizing by-products from fruit and vegetable processing is a crucial and challenging task globally (Homayouni *et al.*, 2013: Tran, 2006). Pineapple pomace contains high levels of dietary fiber, vitamins, and minerals (Wang *et al.*, 2002) Dietary fiber is mainly incorporated into cereal-based products, particularly bakery products.

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fruit and vegetable wastes in food products for human consumption (Nagarajaiah and Prakash, 2016). Studies have explored the incorporation of pineapple pomace in sausages and as fiber enrichment in extruded products (Selani et al., 2014). Orange peel powder has been utilized as a source of dietary fiber in cake production (Montalvo-González et al., 2018); Zaker et al., 2017). Developing beef patties using pumpkin pulp has been researched and preparing high-fiber biscuits from Chinese olive pomace has been reported (Serdaroğlu et al., 2018: Lin et al., 2017). Fiber-enriched wheat bread has also been developed (Chareonthaikij et al., 2016). Studies have indicated that adding pineapple pomace to the diet of diabetic rats improved glucose metabolism and reduced inflammation (Hou et al., 2020). Another study found that pineapple pomace may help reduce cholesterol levels and improve liver function in mice (Yun et al., 2021).

This study aims to utilize pineapple and orange pomace to Yoghurt was produced according to the method of Awolu and enhance yogurt, improving its physicochemical, nutritional, Olofinale (2016) with slight modifications. Five samples of and functional properties.

### **Materials and Methods**

Skimmed milk and the starter culture were purchased from a Supermarket in Akure, Ondo State, Nigeria. Orange and Pineapple fruits were obtained at Oja Oba market, Akure, Ondo State, Nigeria. The fruit samples were authenticated at the Department of Food Science and Technology, Federal University of Technology, Akure, Ondo State, Nigeria. The reagents used for the analyses were of analytical grade.

### **Preparation of Orange Pomace**

Orange (Citrus sinensis) pomaces were prepared in the Food Processing Laboratory, Department of Food Science and

**Table 1:** Formulation of voghurt samples

24010 201 officiation of Jognatic samples					
Labels	Skimmed Yoghurt (%)	Pineapple Pomace (%)	Orange Pomace (%)		
YOA	100	0	0		
YOB	75.04	22.16	2.87		
YOC	84.32	13.65	3.03		
YOD	78.82	16.32	4.88		
YOE	78.05	16.31	5.64		

YOA= 100% Skimmed Yoghurt

YOB=75.04% Skimmed Yoghurt, 22.65% Pineapple Pomace and 2.87% Orange Pomace

YOC= 84.32% Skimmed Yoghurt, 13.65% Pineapple Pomace and 3.03% Orange Pomace

YOD= 78.82% Skimmed Yoghurt, 16.31% Pineapple Pomace and 4.88% Orange Pomace

YOE = 78.0 % Skimmed Yoghurt, 16.31% Pineapple Pomace and 5.64% Orange Pomace

### Determination of pH and Total Titratable Acid of Yoghurt

The pH was measured using a calibrated pH meter (Ashogbon Determination of Free Fatty Acids AOAC (2005).

### **Determination** of Total Soluble Solids **Determination**

The total soluble solids of the yoghurt samples were measured using a digital refractometer (Hanna Instruments 96801 Determination of Viscosity Digital Refractometer, USA) at 25 °C and expressed in °Brix. The measurement of viscosity was conducted using a The sugar content was measured at 20 °C with an eclipse rotational viscometer (RION, Tokyo Japan) according to the refractometer and the result was expressed in °Brix (% sucrose) method of AOAC (2005). 100 mL of sample was loaded into (Zhang and McCarthy, 2013).

2012). Several authors have reported enriching fibers from Technology, Federal University of Technology, Akure, Ondo State, Nigeria. Deseeded orange pomace was oven-dried (Model 320, Gallenkamp, England) at 80 °C for 12 h and subsequently milled (Asiko Attrition Mill, Lagos, Nigeria; Serial No A11). The pomace powder was sieved using a 500 um aperture sieve and packed in an air-tight container until used (Arijit et al., 2020).

### **Preparation of Pineapple Pomace**

The pineapple was washed, peeled, and oven-dried (Model 320, Gallenkamp, England) at 80 °C for 12 h and milled (Asiko Attrition Mill, Lagos, Nigeria; Serial No A11). The pineapple pomace powder was sieved using a 500 µm aperture sieve and packed in an air-tight container for further analysis (Awolu et al., 2019).

### Preparation of Processed Yoghurt made with Pomace

yoghurts were produced using skimmed milk, yoghurt enriched with pineapple, and orange pomace. The Yoghurts were coded YOA, YOB, YOC, YOD and YOE according to their formulation in Table 1. Dried powder milk (250 g) was reconstituted to make up 500 mL using sterile water. The milk was heated to 40 °C. The samples were pasteurized at 90 °C for 15 min. The mixture was cooled to 43 °C and inoculated with a commercial yoghurt starter culture (pure culture of mixed strain of Lactobacillus bulgaricus and Streptococcus thermophillus). The mixture was stirred for complete homogenization. The samples were incubated at about 44 °C for 7 h until curds were formed at pH 4.5. The curds were broken with a hand stirrer to form a smooth homogenous product, and stored in a refrigerator at a temperature of 4 °C for 24 h preparatory for sensory evaluation.

and Akinayo, 2014) and the total titratable acid (TTA) content 95% neutralized isopropanol and phenolphthalein indicator of the yoghurt was determined according to the method of were added to the liquid fat sample. The sample was titrated with NaOH, and each sample's percent free fatty acid was calculated. The color of the sample appeared pink and and Brix persisted for 30 s. The volume of sodium hydroxide used was recorded (Pike and O'Keefe, 2017).

### **Determination of Antioxidant Properties**

The ferric reducing antioxidant property of the extract was determined. 0.25 ML of the sample extract was mixed with 0.25 ML of 200 mM of Sodium phosphate buffer pH 6.6 and 0.25 ML of 1% KFC. The mixture was incubated at 50 °C for 20 min, thereafter 0.25 ML of 10% TCA was also added and centrifuged at 2000 x g force for 10 min. Exactly 1 mL of the supernatant was mixed with 1mL of distilled water and 0.1% FeCl<sub>3</sub>, and the absorbance was measured at 700 nm (Pulido et al., 2000).

(1, 1- diphenyl-2 picryhydrazyl) was determined using the Whatman filter. Exactly 25 mL of the filtrate was later method of Gyamfi et al. (1999). About 1 ML of the extract was transferred into a conical flask and titrated with 0.1m KMnO4 mixed with 1 ML of the 0.4 mM methanolic solution of the at temperatures between 80-90 °C until a pink colour persisted DPPH the mixture was left in the dark for 30 min before for 15 sec was obtained. measuring the absorbance at 516 nm.

The ability of the extract to chelate Fe<sup>2+</sup> was determined using a modified method. Briefly, 150 mM FeSO<sub>4</sub> was added to a reaction- mixture containing 168 mL of 0.1 M Tris-HCl, pH 7.4, 218 mL saline, and extract, and the volume was made up to 1 mL with distilled water. The reaction mixture was incubated for 5 min, before the addition of 13 mL of 1, 10phenantroline; the absorbance was read at 510 nm (Puntel et al., 2005).

The 2, 2'-azino-bis (3-ethylbenthiazoline-6-sulphonic acid) (ABTS) scavenging ability of the extract was determined according to the method described by Re et al., (1999). The ABTS was generated by reacting a 7 mM ABTS aqueous solution with K2S2O8 (2.45 mM/l, final conc.) in the dark for 16 h, while the absorbance at 734 nm was adjusted to 0.700 with ethanol. 0.2 mL of the appropriate dilution of the extract absorbance was read at 732 nm after 15 min.

The TROLOX equivalent antioxidant capacity subsequently calculated. The total phenolic content was measured using the Folin-Ciocalteu reagent and was reported in milligrams of Gallic Acid Equivalents (GAE) per 100 gm (Zheng and Wang, 2001).

# Determination of Antinutritional Qualities of Yoghurt **Samples**

Tannin determination: 0.2 g of finely ground sample was weighed into a 50 mL sample bottle, and 10 mL of 70% aqueous acetone was added and properly covered. The bottle was put in an ice bath shaker and shaken for 2 h at 30 °C. Each solution was then centrifuged and the supernatant was stored in ice. Standard tannin acid solutions were prepared. Half a mL (0.5 mL) of Folin ciocateau reagent was added into the sample. The standard aliquots and 2.5 mL of 20% Na<sub>2</sub>CO<sub>3</sub> was added into the solution. The votexed solution was incubated at room temperature for 40 min, and the absorbance was read at 725 nm against a reagent blank concentration of the same solution from a standard tannic (Makkar and Goodchild. 1996).

the test cell. The rotor was dipped into the sample and allowed Phytate was determined by the method of Wheeler and Ferrel's to spin until the needle scale pointer stopped at a certain scale. (1971). The samples were soaked for three hours in a solution of 100 mL 2% HCl and then filtered through a No 1 Whatman filter paper. From the filtrate, 25 mL was taken and poured into a conical flask, and 5 mL was added to 0.3% ammonium thiocyanate solution which served as an indicator. The required acidity was achieved by blending 53.5% distilled water with the solution. This mixture was titrated against 0.00566 g/ml of iron (iii) chloride solution containing 0.00195 g of iron per milliliter until a brownish-yellow colour which remained constant for 5 mins was formed.

The oxalate content was determined using the method of Baker (1952). One gram (1 g) of the sample was soaked in 75 mL of The free radical scavenging ability of the extract against DPPH 1.5 N H<sub>2</sub>SO<sub>4</sub> solution for an hour, and filtered through a No 1

> The spectrophotometric method of Brunner (1994) was used to determine the saponin content. About 2 g of the sample was weighed into a 250 mL beaker while 100 mL of Isobutyl alcohol was added. A shaker was used to shake the mixture for 5 h to ensure uniform mixing. The mixture was filtered with No 1 Whatman filter paper into a 100 mL beaker containing 20 mL of 40% saturated magnesium carbonate solution (MgCo3). The mixture obtained was filtered through No 1 Whatman filter paper to get a clean colourless solution. It was allowed to stand for 30 min for the colour to develop. The absorbance was read against the blank at 380nm.

Five gram (5g) of the sample was weighed into a 250 mL beaker and 200 mL of 10% acetic acid in ethanol was added and allowed to stand for 4 min to determine alkaloid. This was filtered and the extract was concentrated in a water bath to onequarter of the original volume. Concentrated ammonium was then added to 2.0 ML of ABTS solution and the hydroxide was added dropwise to the extract until the precipitation was complete. The whole solution was allowed to settle, and the precipitate was collected, washed with dilute ammonium hydroxide, and filtered. The residue (alkaloid) was dried and weighed (Harbone, 1973).

### **Statistical Analysis**

Statistical Package for Social Science version 21 (IBM SPSS Statistics, US) was used to examine the results statistically. The mean and standard deviation of the samples were determined and a One-Way Analysis of Variance (ANOVA) was done for comparison. However, the statistically significant difference of all samples analyzed was performed  $p \le 0.05$  (SPSS, 2011).

## **Results and Discussion**

# Physicochemical Composition of the Yoghurt

All samples showed an acidic pH with a range of 3.93 to 4.50 as shown in Table 2 below. The acidic pH range observed in the samples (3.94 - 4.22) aligns with the report of Arijit et al., (2020), that the acidity of yoghurt increased with an increase in the concentration of the orange pomace powder. It is believed that the fermentation process and the actions of lactic acidity (Aderinola and Adeniran, 2015).

When supplemented with orange and pineapple pomace, the acidity of the yogurt was further enhanced. The lactic acid bacteria were found to be responsible for converting lactose in the yogurt samples into lactic acid, thereby increasing the acidity. The pH decrease rate of 3.39% in sample YOB contained a higher percentage of sugar from orange and pineapple, as indicated by the Brix value. Previous studies involving the use of wine grape pomace in yogurt during cold storage indicated that pomace increases the pH value of wine (Tseng and Zhao, 2013). Similar results were also achieved when yogurt was fortified with dried apple peel powder by agreed with FDA specifications for yogurt acidity of 4.6 or culture used, could also increase the fatty acid content in the lower (Srivastava et al., 2015). Samples YOB and YOE, which yogurt. The total titratable acidity of the yogurt in this study contained pomace, exhibited significantly higher Brix values increased with the addition of pomaces. It is believed that the compared to the control yogurt (YOA). Fruits naturally contain fermentation of lactose by lactic acid bacteria, as well as the sugars, such as fructose and glucose, which are released from presence of citric acid and ascorbic acid in orange pomace, the pomace when added to yogurt. As a result, the overall sugar contributed to the increased titratable acidity (Aderinola and content of the yogurt samples.

The viscosity of the yogurts ranged between 3.78 and 4.39 Pa.s. While differences in viscosity may be attributed to the Antinutrient Properties of Yoghurt Made with Orange and type of raw material used, the gradual decline in viscosity in **Pineapple Pomace** yogurts with pomaces could be due to the liquefaction of sugar The result of the antinutrient properties of yoghurt made with during fermentation. Similar trends were reported in yogurt orange and pineapple pomace is presented in Table 3.

acid bacteria influence the pH levels, resulting in improved fortified with dried apple peel powder (Zhou 2018). The total soluble solid (TSS) results revealed that sample YOB had the highest solubility. Generally, adding pomace to the samples conferred higher TSS values than the control yogurt (YOA). The total solids content in the yogurt indicates the amount of sugar and ash present. The fermentation process of lactic acid bacteria resulted in the production of metabolites in the form of lactic acid, thus contributing to the total dissolved solids (Ani et al., 2018). The free fatty acid content in the study ranged from 6.30 to 12.00%. This value was approximately two times higher than in a previous study (Noreen et al., 2019). It is believed that the high level of acetic acid in the present study may have led to this difference. Furthermore, the Zhou (2018). The pH values obtained in the present study metabolism of sugar and amino acids, as well as the type of Adeniran 2015).

**Table 2:** Physicochemical composition of yoghurt yoghurt made with orange and pineapple pomace

Samples	pН	BRIX (%)	TTA (%)	TSS (%)	FFA (%)	Viscosity (Pa.s)
YOA	4.50±0.01a	10.50±0.70°	6.20±82 °	6.00±1.41 <sup>d</sup>	6.30±0.10 °	4.39±0.00 a
YOB	$3.39\pm0.04^{a}$	17.00±0.00 a	$8.50\pm82^{\ b\ c}$	10.50±0.64 a	10.05±63 b	3.78±0.25 °
YOC	$4.40\pm0.13^{a}$	$14.00 \pm 0.00^{b}$	10.99±92 b	8.80±1.13 ab	$9.10\pm0.70^{b}$	$4.16\pm0.06^{ab}$
YOD	4.30±0.13a	11.50±0.70 °	14.93±1.56 a	$6.30\pm0.01^{\text{ c}}$	10.70±4.25 ab	4.03±0.04 bc
YOE	$4.28\pm0.08^{a}$	11.50±0.70°	11.63±1.60 <sup>b</sup>	7.13±2.61 b	12.00±0.56 a	$4.04\pm0.42^{\text{ bc}}$

Results are mean values of duplicate determination ± standard deviation. Mean values within the same column having the same letter are not significantly different at  $p \le 0.05$ .

**Table 3:** Results of anti-nutrient content of yoghurt samples

Samples	Saponins	Oxalate	Phytate (mg/mL)	Tannins	Alkaloids
	(mg/100g)	(mg/100g)		(mg/mL)	(mg/g)
YOA	10.10±0.39 d	0.18±0.00 e	$2.06\pm0.58^{d}$	6.90±0.40 °	5.18±0.50 d
YOB	18.90±0.51 a	1.94±0.63 a	9.50±0.58 a	17.85±1.37 a	6.70±0.35 a
YOC	15.36±0.39 b	1.67±0.63 b	$6.53\pm0.58^{b}$	17.98±0.85 a	$5.61\pm0.50^{b}$
YOD	13.98±0.13 <sup>b</sup>	$0.54\pm0.00^{\circ}$	$4.94\pm0.00^{\text{ c}}$	12.47±0.34 b	5.55±0.01 bc
YOE	11.63±1.02°	$0.40\pm0.06^{\rm d}$	4.72±0.39 °	11.92±2.56 b	5.46±0.01 b

Results are mean values of duplicate determination ± standard deviation. Mean values within the same column having the same letter are not significantly different at  $p \le 0.05$ .

antinutrient content. In our samples, sample YOB, which contained the highest level of pineapple pomace exhibited the highest antinutrient levels. The study found that the oxalate content ranged from 0.18 to 1.94 mg/100g. The significant oxalate content in the samples may be due to the low oxalate content in the animals' food sources. A study by Ani et al., (2018) reported lower oxalate values ranging from 0.16 to 0.62 mg/100g in yogurt prepared from moringa seed milk, soybean, and Bambara nut. The phytic acid content of the yoghurt in this study ranged from 2.06 to 9.50 mg/100g. Interestingly, the phytic acid content is comparable to the 3.28 to 11.70 mg/100g range reported by Zaker et al., (2016), as well as the values reported for yogurt prepared from soybeans, Bambara nut and moringa seed milk (Ani et al.,

The presence of pomaces in the yogurt is crucial for assessing the 2018), which varied from 2.37 to 8.78 mg/100g. Despite being an antinutrient, phytate has beneficial effects, such as lowering blood glucose levels, plasma cholesterol, and triacylglycerol levels (Ogodo et al., 2019). Studies have shown that fermentation reduces the amount of anti-nutritional factors (Etsuyankpa et al., 2015). The saponin content in the study ranged from 10.10 to 18.90 mg/100g higher than the range 0.86 to 3.75 mg/100g saponin content obtained from yogurt made from moringa and bambara nut milk (Ani et al., 2018). The high saponin content observed in our study may be a result of the incorporation of pomaces. Saponins have been reported to have anticancer properties, as well as hypocholesterolemic effects (Man et al., 2010).

# **Pineapple Pomace**

The antioxidant properties of skimmed voghurt with orange and pineapple pomace are presented in Table 4. Probiotics in yogurt, such as Lactobacillus acidophilus and Bifidobacterium lactis enhance antioxidant activities, which can be beneficial for individuals with Type 2 Diabetes Mellitus (Simwaka et al., 2017). Yoghurt samples with pomace exhibited higher overall antioxidant activities compared to the sample without pomace, except for the ABTS test, where there were no significant (p< bioactive compounds (Huang et al. 2011), which could increased antioxidant activities.

Antioxidant Properties of Yoghurt made with Orange and contribute to the high phenolics content observed in samples YOB (0.10 mg GAE/100 g) and YOC (0.11 mg GAE/100 g). It is known that the reducing power of Fe<sup>2+</sup> is associated with antioxidant activity, a relationship that has been established with numerous bioactive compounds isolated from natural plants (Siddhurajua et al., 2017). There was a significant increase in the Fe<sup>2+</sup> levels in yogurt containing pineapple and orange pomace, ranging from 21.75-29.62, compared to 16.69 in 100% skimmed yogurt without pomace. The high antioxidant activity is attributed to the radical absorbing effect 0.05) differences among all the samples, including those with of the bioactive compounds in the enriched yogurt. Pineapple pomace. Additionally, sample YOB, which had the highest pomace has also been shown to have antioxidant properties pineapple pomace incorporation, showed the best overall and may help protect against oxidative stress (Hilkal et al., antioxidant activities. Pineapples are known to contain various 2021). In general, adding blended pomace to yogurt led to

**Table 4:** Antioxidant properties of the yoghurt

Samples	Phenol	FRAP	DPPH	ABTS	$Fe^{2+}$
	(mg GAE/g)	(mg/g)	(%)	(mMol/g)	
YOA	0.07±0.00°	8.84±0.73 d	57.26±0.36°	0.01±0.00a	16.69±0.84°
YOB	$0.10\pm0.02^{ab}$	18.75±0.80 a	$68.26\pm0.25^{a}$	$0.02\pm0.00^{a}$	$29.62\pm2.42^{a}$
YOC	$0.11\pm0.00^{a}$	17.75±1.35 b	$61.86 \pm 0.36^{ab}$	$0.01\pm0.00^{a}$	$24.40\pm2.54^{ab}$
YOD	$0.08\pm0.02^{c}$	12.30±0.36 °	$59.61\pm0.10^{b}$	$0.01\pm0.00^{a}$	$21.75\pm2.67^{bc}$
YOE	$0.08\pm0.00^{c}$	12.14±0.18 °	$59.55 \pm 0.88^{b}$	$0.02\pm0.00^{a}$	$22.52\pm1.09^{b}$

Results are mean values of duplicate determination ± standard deviation. Mean values within the same column having the same letter are not significantly different at p≤0.05.

### Conclusion

The pineapple and orange pomace affect the physicochemical properties of the yoghurt by reducing pH and Viscosity while the brix, total titratable acidity, total soluble solids, and free fatty acids were increased. The yoghurts fortified with pomace had a positive effect on nutritional properties compared to the control but sample YOB showed the strongest desirable antinutrient and antioxidant properties. The addition of pineapple and orange pomace to yoghurt reduces its viscosity. Therefore, orange and pineapple pomace can serve as an industrial by product in yoghurt and can successfully be utilized for the development of fiber-enriched yoghurt with improved health benefits.

### **Declarations**

### **Competing Interest**

The authors declare no competing interest.

### **Authors' Contributions**

All listed authors contributed equally to the research process, literature writing, review and editing of the article.

### References

Aderinola, T.A. & (in full) Adeniran, A.E. (2015). Effects of storage on physicochemical properties of orange-watermelon juice. Ann. Technology, Science. 16. 326-332. https://www.researchgate.net/publication/311435639.

Ani C., Amove J. & Igbabul O., (2018). Physicochemical, microbiological, and sensory properties and storage stability of plant-based yoghurt produced from Bambara nut, soybeans, and moringa seed milk. American Journal of Food and Nutrition; 6(4):115-125. doi: 10.12691/ajfn-6-4-4 AOAC, (2005).

Official Methods of Analysis, Association of Official Analytical Chemists, 15th Edition. Horwitz, W. and Latimer, G.W. (Ed). AOAC International, Maryland-USA.

Arijit, M., Maity, S., Roy, S., Ghosh, B. C. & Kundu, S. (2020). Utilization of pomace from orange juice production in yoghurt as a value-added ingredient. Journal of Food Processing and Preservation, 44(7), e14574. https://doi.org/10.1111/jfpp.14574.

Asoghon A. O & Akintayo E. T., (2014). A recent trend in the physical and chemical modification of starches from different botanical http://dx.doi.org/10.1002/star.201300106 Starch/Starke. 66:41-57.

Awolu, O. O., Magoh, A. O., & Ojewumi, M. E. (2020). Development and evaluation of extruded ready-to-eat snack from optimized rice, kersting's groundnut, and lemon pomace composite flours. Journal of food science and technology, 57, 86-95. doi: 10.1007/s13197-019-04033-9.

Awolu, O. O., Osigwe, M. A. & Ifesan B. O. T. (2019). Nutritional and antioxidant potential of rice flour enriched with Kersting's groundnut (Kerstingiella geocarpa) and lemon pomace. International Journal of Food Studies, 53(2):1151-63. doi: 10.1007/s13197-015 2121-8

Awolu, O. O., Osemeke, R. O., & Ifesan, B. O. T. (2016). Antioxidant, functional, and rheological properties of optimized composite flour, consisting of wheat and amaranth seed, brewers' spent grain, and apple pomace. Journal of food science and technology, 53(2):1151-63. doi: 10.1007/s13197-015-2121-8.

Awolu, O.O and Olofinale, S.J (2016). Physico-chemical, Functional, and pasting properties of native and chemically modified water yam (Discorea alata) starch and production of water yam based Starch/Starke. 68:18 yoghurt. DOI:10.13140/RG.2.2.33285.45286.

Brunner, J.H. (1984): Direct spectrophotometer determination of saponin. Animal Chemistry 34: 1314-1326.

Baker C.J.A (1952). The determination of oxalates in fresh plant material. Analyst, 916: 304-309, 10.1039/an9527700340.

Chareonthaikij, P., Uan-On, T., & Prinyawiwatkul, W. (2016). Effects of pineapple pomace fibre on physicochemical properties of composite flour and dough, and consumer acceptance of fibreenriched wheat bread. International Journal of Food Science & 1120 -1129. https://doi.org/10.1111/ijfs.13072

- Dhingra, D., Michael, M., Rajput, H., & Patil, R. T. (2012). Dietary fibre in foods: A review. Journal of Food Science and Technology, 49(3), 255- 266. https://doi.org/10.1007/s13197 011-0365-5
- Etsuyankpa, M. B., Gimba, C. E., Agbaji, E. B., Omoniyi, I., Ndamitso, M. M., & Mathew, J.T., (2015). Assessment of the Pike, O. A., & O'Keefe, S. (2017). Fat Characterization. Food effects of microbial fermentation on selected anti-nutrients in the products of four local cassava varieties from Niger state, Nigeria. American Journal of Food Science and Technology, 3(3), 89-96. Pulido R, Bravo L, Saura-Calixto F (2000) Antioxidant activity of doi: 10.12691/ajfst-3-3-6.
- Gyamfi, M.A., Yonamine, M. & Aaniya, Y., (1999): Free radical scavenging action of medicinal herbs from Ghana: thonningia sanguine on experimentally induced liver injuries. General Pharmacology. 32: 661 – 667. <a href="https://doi.org/10.1016/S0306-">https://doi.org/10.1016/S0306-</a> 3623(98)00238-9.
- Harbone, J. B. (1973). Phytochemical methods. Chapman and Hall Re R., Pellegrin N., Proteggente A., Pannala A., Yang M. & Rice-London, 10-2.
- Hikal W. M., Mahmoud A. A, Hussein A. H. Said-Al Ahl, Amra Bratovcic, Kirill G. Tkachenko, Miroslava Kačániová & Ronald Maldonado Rodriguez (2021). Pineapple (ananas comosus 1. merr.), waste streams, characterisation and valorisation: an overview. Open Journal of Ecology, 11, 610-634. DOI: 10.4236/oje.2021.119039.
- Homayouni, A., Azizi, A., & Ehsani, M. R. (2013). Citrus byproducts as a source of functional and nutritional ingredients. Innovative Food Science & Emerging Technologies, 20, 1-10. doi: 10.1016/j.ifset.2013.07.011.
- Hou, J., Wang, Y., He, J., Zhang, H., Li, C., & Yang, G. (2020). Pineapple pomace improves glucose metabolism in diabetic rats by modulating gut microbiota. Food Research International, 137,
- physicochemical properties of fiber-rich fraction from pineapple peels as a potential ingredient. Journal of Food and Drug Analysis, 19(3), 4.
- Hussein, A. M., Kamil, M. M., Hegazy, N. A., Mahmoud, K. F., & Ibrahim, M. A. (2015). Utilization of some fruits and vegetables by-products to produce high dietary fiber jam. Food Science and Quality Management, 37, 39-45.
- Lin, S., Chi, W., Hu, J., Pan, Q., Zheng, B., & Zeng, S. (2017). Sensory and nutritional properties of Chinese olive pomace based high fibre biscuit. Emirates Journal of Food and Agriculture, 39(7), 495–501. https://doi.org/10.9755/ejfa.2016-12-1908.
- Man S., Gzo W., Zhang Y., Liu C. (2010). Chemical study and medical application of saponnins as anti canceragent. Fitoterapia. 1(7):703-14. doi: 10.1016/j.fitote.2010.06.004.
- Markkar, A. O. S. & Goodchild, A. V. (1996); Qualification of Siddhurajua P., Jent K., & Joy (2017). Studies on nutritional profile Tannis. A laboratory Manual.International Centre of Agricultural Research in Dry Areas (ICRDA). Alleppo Syria, IV. 25pp
- Montalvo-González, E., Aguilar-Hernández, G., Hernández-Cázares, A.S., Ruiz-López, I. I., Pérez-Silva, A., Hernández-Torres, J., & Vivar-Vera, M. D. L. Á. (2018). Production, chemical, physical and technological properties of antioxidant dietary fiber from pineapple pomace and effect as ingredient in sausages. CyTA-Journal of Food, https://doi.org/10.1080/19476337.2018.1465125
- Nagarajaiah, S. B., and Prakash, J. (2016). Chemical composition and bioactivity of pomace from selected fruits. International Journal https://doi.org/10.1080/15538 362.2016.1143433, of Fruit Science, 16(4), 423–443.
- Noreen S., Faiza I., Manzoor M. F., Anjum F. M., (2019). Pineapple waste: A review of its potential health benefit...Journal of Food Science and Technology. 56(5):2429-2436.
- Ogodo, A. C., Agwaranze, D. I., Aliba, N. V., Kalu, A. C., & Nwaneri, C. B. (2019). Fermentation by lactic acid Bacteria consortium and its effect on antinutritional factors in maize flour. Biological Journal of Sciences. 19. 17-23. DOI:10.3923/jbs.2019.17.23.

- Olabiran, T. E., Awolu, O. O., & Ayo-Omogie, H. N. (2023). Quality chracterization of functional soy-based yoghurt incorporated with scent leaf (Ocimum gratissimum) essential oil microcapsules. Food Chemistry Advances, 100336. DOI:10.1016/j.focha.2023.100336.
- Science Text Series, 407-429. https://doi.org/10.1007/978-3-319-45776-5\_23.
- dietary polyphenols as determined by a modifed ferric reducing/ antioxidant power assay. Journal of Agriculture and Food Chemistry 48(8):3396-3402. DOI: 10.1021/jf9913458.
- Puntel, R.L., Nogueira C. W. & Rocha, J. T. (2005). Krebs cycle intermediates modulate Thiobarbituric. Neurochemical Research ; 30(2):225-35. doi: 10.1007/s11064-004-2445-7
- Evans C., (1999) Antioxidant activity applying an improve ABTS Radication decolourization assay. Free Radical Biology and Medicine. 26: 1231-1237. doi: 10.1016/s0891-5849(98)00315-3.
- Rodríguez, G., Lemes, A., & Lomónaco, M. (2006). Dietary fiber from fruit by-products enhances the water-holding capacity and other functional properties of meat products. Meat Science, 73(3), 405-413.
- Sabanis D., Lebesi D., & Tzoa C., (2009). Development of fibre enriched gluten-free bread. A response surface methodology study. International Journal Food Sciences and Nutrition. 60(4):174-190. DOI:10. 1080/09637480902721196.
- Sandez T. Lopez-Nicholaz R. & Planes D (2017). In-vitro modulation of gut microbiota by whey protein to preserve intestinal health. Food and Function: 8, 3053-3063. https://doi.org/10.1 039/C7FO00197E.
- Huang, Y. L., Chow, C. J., & Fang, Y. J. (2011). Preparation and Selani M. M., Brazaca C. G., Dias S. D., Ratnayake W. S. & Flores R. A.(2014) Characterisation and potential application of pineapple pomace in anm extruded product for fiber enhancement. Food Chemistry.163:23-30.http://doi.org/10.1016/j.foodchem2014.04.076/
  - Serdaroglu M., Kavasan H.S., Ipek G., & Ozturk B., (2018). Evaluation of the quality of beef patties with dried pumpkinpulp and seed. Korean Journal of Food Science of Animal Resource. 38(1):1-13. DOI:10.5851/kosfa.2018.38.1.001.
  - Simwaka, J. E., Chamba, M. V. M., Huiming, Z., Masamba, K. G., & Luo, Y. (2017). Effect of fermentation on physicochemical and antinutritional factors of complementary foods from millet, sorghum, pumpkin and amaranth seed flours. International Food Research Journal, 24(5), 18691879. https://www.researchgate.net/publication/322887426.
  - and antioxidant potential of different Discorea spp with Pelecthrathus rotundifolus. International Journal Current 65-74. Pharmaceutical Research, 9(4) DOI:10.22159/ijcpr.2017v9i4.20959
  - SS. Statistical packages for the social sciences, Version 20.0 IBM Corp., Armonk, NY, USA; 2011.
  - Srivastava, P., Prasad, S. G. M., Ali, M. N., & Prasad, M. (2015). Analysis of a8-ntioxidant activity of herbal yoghurt prepared from different milk. The Pharma innovation Journal, 4(3): 18-20.https://www.researchgate.net/publication/292129898\_Analys is\_of\_antioxidant\_activity\_of\_herbal\_yoghurt\_prepared\_from\_d
  - Tran, A.V. (2006). Chemical analysis and pulping study of pineapple crown leaves. Industrial Crops and Products, 24, 66-74. DOI:10.1016/j.indcrop.2006.03.003.
  - Tseng A, & Zhhao Y., (2013). Wine grape promace on antioxidant, dietary fibre for enhancing nutritional value and improving the storability of yoghurt and salad dressing. Food Chemistry. 138(1): 356-365. DOI:10.1016/J. Foodchem.
  - Wang, J., Rosell, C.M. & Benedito de Barber, C. (2002). Effect of the addition of different fibres on wheat dough performance and

- bread quality. Food Chemistry, 79, 221–226. DOI: 10.1016/S0308-8146(02)00135-8
- Wheeler, E.L. and R.A. Ferrel, 1971. A method for phytic acid determination in wheat and wheat flour.Cereal Chemistry, 48:313-314. cerealsgrains.org/publications/cc/backissues/1971/Do cuments/chem48\_312.pdf.
- Yun, J. W., Lee, D. W., Park, E. H., Lee, J. H., Kim, D. H., Lee, S. H., & Kim, K. S. (2021). Pineapple pomace ameliorates hypercholesterolemia and hepatic steatosis in high-fat diet fed mice. Journal of Agricultural and Food Chemistry, 69(18), 5303-5311.
- Zaker, M. A., Sawate, A. R., Patil, B. M., Sadawarte, S. K., & Kshirsagar, R. B. (2017). Utilization of orange (Citrus sinesis) peel powder as a source of dietary fibre and its effect on the cake

- quality attributes. International Journal of Agricultural Sciences, 13(1), 56–61. <a href="https://doi.org/10.15740/HAS/IJAS/13.1/56-61">https://doi.org/10.15740/HAS/IJAS/13.1/56-61</a>.
- Zhang, L., McCarthy, M.J., 2013. Assessment of pomegranate postharvest quality using nuclear magnetic resonance. Postharvest DOI: 10.1016/j.postharvbio.2012.11.006 Biology and Technology. 77, 59–68.
- Zheng, W. and S.Y.Wang, 2001. Antioxidant activity and phenolic compounds in selected herbs, Journal of Agricultural Food Chemistry. 49:5165-5170. DOI: 10.1021/jf010697n.
- Zhou Ze Hu (2018) .Effect of dried apple peel powder on the rheological and sensory properties of drinking yoghurt. Agricultural and Food Sciences. @inprocee dings,url={https://api.semanticscholar.org/CorpusID: 103251670."



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